

Evaluating the radioprotective effect of arbutin on mice exposed to megavoltage X-rays based on hematological parameters and lymphocytes micronucleus assay

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ABSTRACT

Background: X-irradiations induce damages to the hematopoietic system by reducing the production of blood cells in bone marrow. In this study, the radioprotective effect of arbutin was investigated in megavoltage x-irradiated mice by measuring changes in hematological parameters and lymphocyte cells with micronucleus assay. **Material and Methods:** Sixty NMRI mice were irradiated with 6 MV photon beam (2 and 4 Gy in one fraction). Various concentrations of arbutin (50, 100, and 200 mg/kg) were injected intra-peritoneal, 2 hours before whole body X-irradiation. Samples of peripheral blood cells were collected from the left ventricle. The frequency of micronuclei in 1000 cells were measured for each sample and level of peripheral blood cells were analyzed. The data were statistically evaluated using one-way ANOVA, and Tukey HSD test. **Results:** X-radiations significantly decreased the hematological parameters such as white blood cells (WBC), lymphocyte (LYM), red blood cells (RBC) counts, and hemoglobin (HGB) levels compared to the control group ($P < 0.001$). The frequency of micronuclei in "2 and 4 Gy X-irradiation + distilled water" groups was significantly higher than "2 and 4 Gy irradiation + 50 mg/kg arbutin", "2 and 4 Gy irradiation + 100 mg/kg arbutin", and "2 and 4 Gy irradiation + 200 mg/kg arbutin" groups, followed by the above-mentioned blood cell parameters were dropped remarkably. **Conclusion:** The present investigation showed that arbutin is a strong radioprotector. There were not any significant differences between the various concentrations of arbutin, however, the concentration of 50 mg/kg showed higher radioprotective effect.

Keywords: Radiation, arbutin, hematopoietic, radioprotector, micronucleus assay.

INTRODUCTION

Ionizing radiations such as X-rays, nuclear

medicine, and radiotherapy are used for medical diagnosis and treatment ^(1,2). But their use is limited because of the undesirable and side

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effects such as normal tissue injury^(3,4).

Radiotherapy is used for the treatment of around half of patients due to the long-term advantages^(2,5). On the other side, this kind of treatment can induce primary lesions in the biomolecular structures and carcinogenic effects on normal tissues⁽⁶⁾, as well as damage to the hematopoietic, gastrointestinal, and central nervous systems⁽⁷⁾.

The effect can be due to the production of free radicals that destroy the vital macromolecules such as proteins, lipids, RNA, and DNA damage/chromosomal mutation, and after exposure, a hematologic crisis occurs, characterized by hypoplasia/aplasia of the bone marrow^(8,9).

Two of the main strategies, non-pharmacological and pharmaceutical, are able to reduce the radiation damages with repairing and recovery enhancers and modifiers of immune system⁽¹⁰⁾. The non-pharmacological strategy can reduce radiation injuries, however, the probability of secondary cancers is still considerably high⁽¹¹⁾. Therefore, focus on radiation protection effects of pharmaceutical products can be helpful against damage when given prior to radiation exposure^(12,13).

There are some plants and herbs beside synthetic antioxidants which protect against radiation injuries⁽¹⁴⁾, for example lettuce oil, aged garlic, and grape seed⁽¹⁵⁻¹⁷⁾. In a previous study, Nadi *et al.*⁽¹⁸⁾ showed the protective effect of arbutin in mice bones as a radioprotector using erythrocytes micronucleus assay. Arbutin is one of the neutral radioprotectors and overcomes the toxic effect of radiation in cancer patients undergoing radiotherapy that has been approved in previous studies⁽¹⁸⁻²⁰⁾. This radioprotector is a naturally occurring glucoside of hydroquinone family and has been traditionally used to treat pigmentary disorders⁽²¹⁾. It is found in a variety of plants including those from the Ericaceae and Compositae families⁽²²⁾. The molecular weight and maximum absorbance spectrum of arbutin are 272.25 g/mol and 267 nm.

Since the hematopoietic system is a radiosensitive tissues, it could be considered as an appropriate indicator of radiation injuries.

One of the best method for measuring chromosome loss and chromosome breakage is micronucleus assay⁽²³⁾. Cytokinesis-block micronucleus (CBMN) assay, which blocked by cytochalasin-B, allows better precision for cell division assessment⁽²⁴⁾.

The radioprotective effect of arbutin on the hematological system have not been reported in previous studies, furthermore, regarding the high radiosensitivity of this system, it will be a good idea to evaluate the radioprotective effect of arbutin based on measurement of hematological parameters. Thus, the aim of this study was to evaluate the biocompatibility of the arbutin as a radioprotector agent in megavoltage therapeutic whole-body x-irradiated mice on hematological parameters at three different concentrations and also lymphocyte cells were assessed utilizing micronucleus assay.

MATERIALS AND METHODS

Animals

Our study was done following Animal Experimentation Ethics Committee and National Research Ethics Board approval with the registration number of "MUBABOL.REC.1391.4" at "22.12.97".

Sixty male NMRI mice, with average age (6-7 weeks) and body weight (25 ± 5 gr) were obtained from animal lab of Iran University of Medical Science (Tehran, Iran). The mice were kept at proper temperature, relative humidity, and light regime (12 hrs. light/12 hrs. dark). They were given fresh water and standard food of a commercial balanced diet.

Irradiation

The mice were placed in a Plexiglas collective cage for whole body exposure by a linear Elekta Compact accelerator (Elekta AB, Stockholm, Sweden) using 6 MV photons. The source-to-skin distance and field size were chosen 100 cm and 20×20 cm², in that order. Two doses of 2 Gy and 4 Gy were applied as single fraction at a dose rate of 200 cGy/min. All irradiations were performed 2 hours after the injection of arbutin. After irradiation the mice were brought back to

the animal lab for the follow up tests.

Chemical

Pure (98%) arbutin powder (Sigma-Aldrich, USA) was injected intra-peritoneal at the concentrations of 50, 100, and 200 mg/kg, 2 hours before whole body x-irradiation. For each mouse, 0.5 cc distilled water was used as solvent.

Experimental design

Two weeks after acclimatization and conditioning, the animals were divided into twelve groups (five mice for each group) in separate Plexiglas cages as following:

- I. "Sham irradiation + distilled water"
- II. "2 Gy X irradiation + distilled water"
- III. "4 Gy X irradiation + distilled water"
- IV. "Sham irradiation + 50 mg/kg arbutin"
- V. "Sham irradiation + 100 mg/kg arbutin"
- VI. "Sham irradiation + 200 mg/kg arbutin"
- VII. "2 Gy X irradiation + 50 mg/kg arbutin"
- VIII. "2 Gy X irradiation + 100 mg/kg arbutin"
- IX. "2 Gy X irradiation + 200 mg/kg arbutin"
- X. "4 Gy X irradiation + 50 mg/kg arbutin"
- XI. "4 Gy X irradiation + 100 mg/kg arbutin"
- XII. "4 Gy X irradiation + 200 mg/kg arbutin"

Sample collection and hematological studies

After 36 hours of irradiation, blood samples were collected from the left ventricle and immediately placed in two tubes. One part was selected to show chromosomal damages of peripheral blood lymphocyte which measured by micronucleus assay and another part was chosen to measure the blood parameter levels.

Complete blood count (CBC) measurement

The CBC was done in differential analysis by veterinary department of Iran University of Medical Sciences. A hematological auto-analyzer (Orphee Mythic 22 Hematological Analyzer; Diamond Diagnostic; USA) was used for measuring CBC.

Micronucleus assay

The heparinized blood samples were added to 4.5 ml complete medium (RPMI 1640

medium, Sigma-Aldrich, USA) containing 20% fetal calf serum, 100 µl/ml phytohemagglutinin (Sigma-Aldrich, USA), 100 IU/ml penicillin (Sigma-Aldrich, USA), 100 µg/ml streptomycin (Sigma-Aldrich, USA) and 2 mM glutamine (Sigma-Aldrich, USA). All cultures were incubated in a proportion condition ($37 \pm 1^\circ\text{C}$ temperature and in a humidity atmosphere with 5% CO₂ and 95% O₂). At 21 h medium was replaced with fresh medium lacking mitogen but containing 4 µg/ml Cytochalasin-B (Sigma-Aldrich, USA) was added to the cultures, and cells were collected by centrifuging (Corning LSE compact centrifuge, CLS6758 model, Sigma-Aldrich, USA; 800 rpm, 5 minutes) at 52 h of incubation ⁽²⁵⁾.

The collected cells were suspended in 0.075 M cold potassium chloride (Merck, Germany) and centrifuged (Corning LSE compact centrifuge, CLS6758 model, Sigma-Aldrich, USA; 800 rpm, 6 minutes) and immediately fixed in a fixative solution (methanol: acetic acid, 6:1; Sigma-Aldrich, USA) for 3 times. Then, the fixed cells were dripped onto clear microscopic slides, air-dried and stained with Giemsa solution (Merck, Germany). Measurement of slides was done ($\times 1000$ magnification with Radical-614 optical microscope model, Radical, USA) to delineate the numbers of micronuclei in cytokinesis-blocked binucleate cells with the cytoplasm remaining intact. Diameter ranging from 1/16 to 1/3 of the diameter of the main nuclei, being non-refractive, no link or overlap with the main nuclei were considered to score small nuclei. For each blood sampling group (irradiated, control, irradiation plus arbutin sample) 1000 cells were assessed to score the frequency of micronuclei.

Statistical analysis

The average counts of WBC, RBC, LYM, and HGB levels and the frequency of micronuclei were compared utilizing one-way analysis of variance (ANOVA), and Tukey's HSD test as the post hoc between the experimental groups to determine which groups were significantly different at 0.05 level. Statistical analysis was carried out through SPSS 16 (SPSS Inc., Chicago, Illinois, US).

RESULTS

Blood cell parameters

The results of the “sham irradiation + distilled water” group did not show significant differences of blood constituents compared to the “sham irradiation + 50 mg/kg arbutin”, “Sham irradiation + 100 mg/kg arbutin”, and “sham irradiation + 200 mg/kg arbutin” groups ($P > 0.05$) (table 1).

Table 1 shows that a remarkable reduction in RBC, WBC, LYM, and HGB blood cell parameters was observed in the “2 and 4 Gy x irradiation + distilled water” groups in comparison with the “sham irradiation + distilled water” group ($P < 0.001$). Furthermore, a significant variation was found in the “2 and 4 Gy irradiation + distilled water” groups compared to the “2 and 4 Gy irradiation + 50 mg/kg arbutin”, “2 and 4 Gy irradiation + 100 mg/kg arbutin”, and “2 and 4 Gy irradiation + 200 mg/kg arbutin” (figures 1 and 2).

Statistical analysis showed hematological factors in the “2 Gy irradiation + 50 mg/kg arbutin” group were considerably higher than the “2 Gy irradiation + 100 mg/kg arbutin” and “2 Gy irradiation + 200 mg/kg arbutin” groups. There was no significant difference in the “2 Gy irradiation + 100 mg/kg arbutin” compared to the “2 Gy irradiation + 200 mg/kg arbutin” group ($P > 0.05$) (table 1).

According to table 1, no significant differences were observed in the levels of the blood cells in the “4 Gy irradiation + 50 mg/kg arbutin” and “4 Gy irradiation + 100 mg/kg

arbutin” groups, but there was a remarkable difference in the “4 Gy irradiation + 200 mg/kg arbutin” compared to the “4 Gy irradiation + 50 mg/kg arbutin” and “4 Gy irradiation + 100 mg/kg arbutin” groups (hematological factors in the “4 Gy irradiation + 200 mg/kg arbutin” were low).

Micronuclei

According to figure 3, there was no side effects after injection arbutin 50, 100, and 200 mg/kg ($P > 0.05$), but a significant increase was observed for the incidence of micronuclei in irradiated group (2 and 4 Gy) compared to the control group ($P < 0.001$).

The frequency of micronuclei found in the “2 and 4 Gy x irradiation + distilled water” groups was significantly higher than the “2 and 4 Gy irradiation + 50 mg/kg arbutin”, “2 and 4 Gy irradiation + 100 mg/kg arbutin”, and “2 and 4 Gy irradiation + 200 mg/kg arbutin” groups ($P < 0.001$).

The comparison into the groups indicated that the frequency of micronuclei in the “2 Gy irradiation + 50 mg/kg arbutin” group was considerably lower than the “2 Gy irradiation + 100 mg/kg arbutin”, and “2 Gy irradiation + 200 mg/kg arbutin” groups, but there were not any differences between the “2 Gy irradiation + 100 mg/kg arbutin”, and “2 Gy irradiation + 200 mg/kg arbutin” groups. No significant differences were observed among the “4 Gy irradiation + 50 mg/kg arbutin”, “4 Gy irradiation + 100 mg/kg arbutin”, and “4 Gy irradiation + 200 mg/kg arbutin” groups.

Table 1. Statistical significance between different groups using Tukey HSD test.

| Blood count | Group I | | Group I | | | Group VII | | Group VIII | Group XII | | Group X |
|----------------------------|----------|-----------|----------|---------|----------|------------|----------|------------|-----------|----------|----------|
| | Group II | Group III | Group IV | Group V | Group VI | Group VIII | Group IX | Group IX | Group X | Group XI | Group XI |
| WBC ($10^9/\mu\text{l}$) | 0 | 0 | 0.0 | 0.057 | 0.061 | 0.039 | 0.045 | 0.055 | 0 | 0.002 | 0.074 |
| LYM ($10^9/\mu\text{l}$) | 0 | 0 | 0.055 | 0.062 | 0.062 | 0.005 | 0 | 0.06 | 0.003 | 0.006 | 0.053 |
| RBC ($10^9/\mu\text{l}$) | 0 | 0 | 0.062 | 0.071 | 0.07 | 0.023 | 0.029 | 0.053 | 0.002 | 0.02 | 0.062 |
| HGB (g/dl) | 0 | 0 | 0.07 | 0.059 | 0.062 | 0.031 | 0.032 | 0.078 | 0.03 | 0.041 | 0.059 |

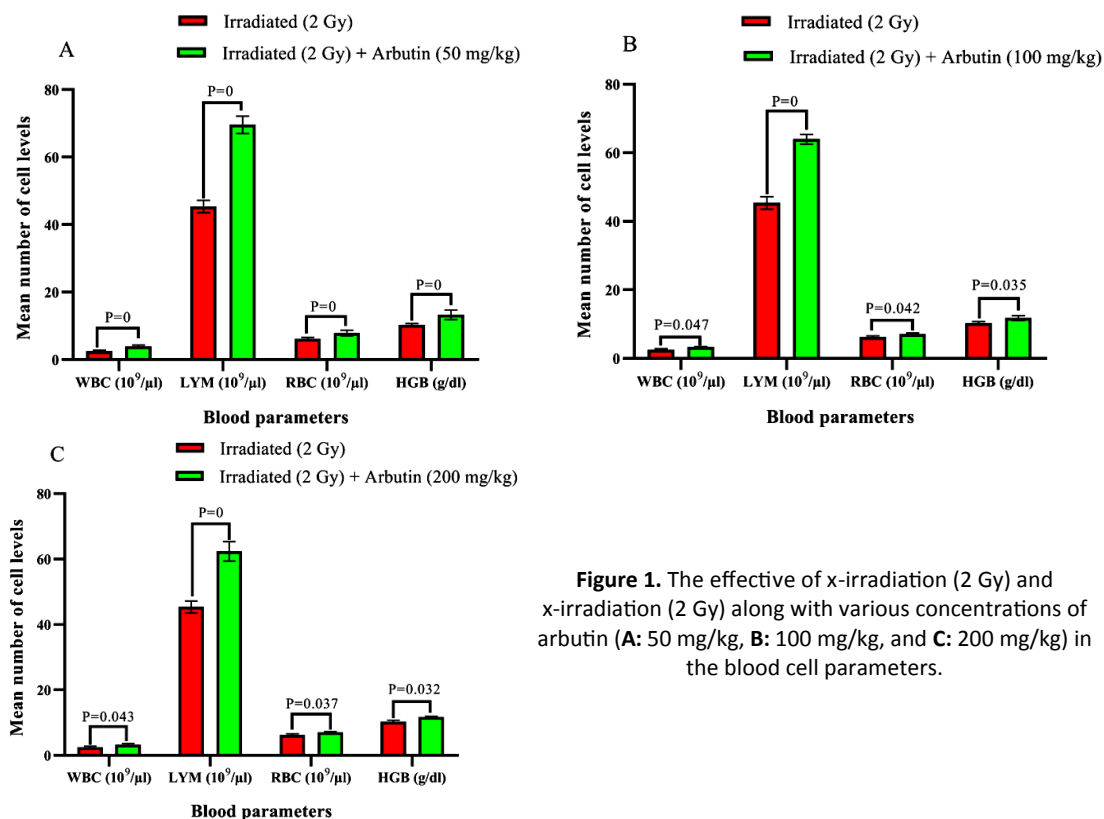


Figure 1. The effective of x-irradiation (2 Gy) and x-irradiation (2 Gy) along with various concentrations of arbutin (A: 50 mg/kg, B: 100 mg/kg, and C: 200 mg/kg) in the blood cell parameters.

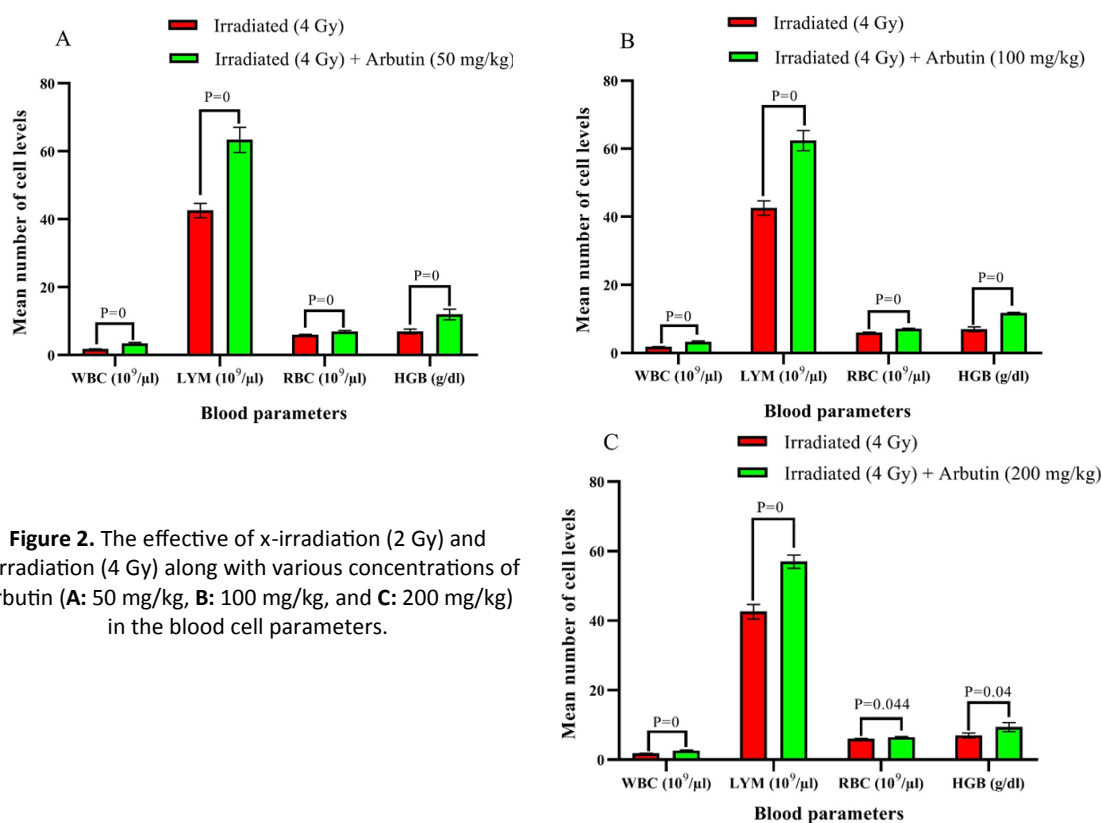


Figure 2. The effective of x-irradiation (2 Gy) and x-irradiation (4 Gy) along with various concentrations of arbutin (A: 50 mg/kg, B: 100 mg/kg, and C: 200 mg/kg) in the blood cell parameters.

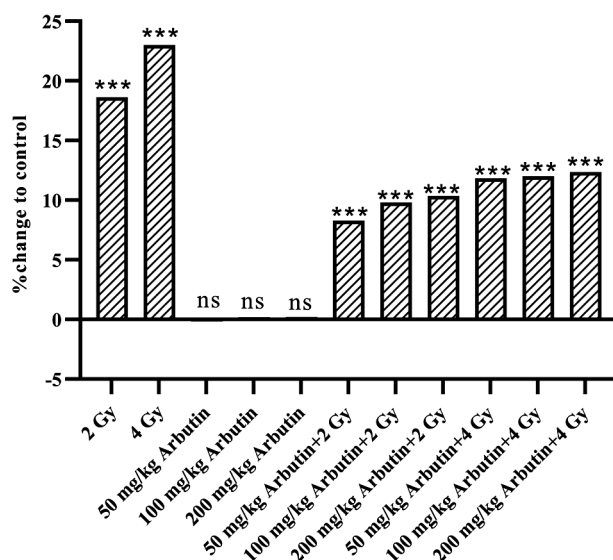


Figure 3. Effects of x-irradiation and different concentrations of arbutin in micronuclei level with respect to the control group.

***Significant difference from the groups compared to the control group at $P < 0.001$.

DISCUSSION

Ionizing radiation such as radiotherapy is one of the important techniques for medical treatment ⁽²⁾, but this technique causes undesirable physiological and biochemical changes in humans and animals ^(26,27). The DNA damage at the chromosome level is an essential part of genetic toxicology. Consequently, this effective of radiation can cause the inhibition in mitosis and decreasing the amount and life span of peripheral blood cells as the most sensitive system ^(15,28,29).

The objective of the current study was to detect what extent of arbutin as a radioprotector can ameliorate the levels of some cellular elements of blood during whole-body x-irradiation on mice. Furthermore, the lymphocyte cell levels were investigated by micronucleus assay.

Our findings showed that x-irradiations (2 and 4 Gy) induced a decrease in blood parameters such as WBC, RBC, LYM, and HGB compared with the control group. The current study corroborates the previous findings of Shaheen and Hassan ⁽³⁰⁾, who declared that x-radiation caused a remarkable decrease in

RBC ⁽³⁰⁾. Also, our results showed that the frequency of the micronuclei in the group which only received radiation (2 and 4Gy) was higher than the control group. Other studies by Hosseinimehr and Nemati ⁽³¹⁾, Shahidi and Mozdarani, ⁽³²⁾ and Shokrzadeh *et al.* ⁽³³⁾ confirmed the current results.

The increasing of micronuclei affected by chromosomal damages induced with radiation. Free radicals like hydroxyl and superoxide radicals produced by radiation can cause the RBC death and also an imbalance between its production and loss. In addition, after irradiation, the count of LYM may be reduced due to the regeneration of intracellular stores of reduced glutathione regarding the oxidative damage. The subsequent decline in the values of WBCs and LYM could be a result of the absence of infection and inflammation, furthermore, the generation of HGB can be reduced because of liver and bone marrow damage resulting from radiation exposure ^(15,17,34,35).

The normal levels of hematological parameters usually measured can be modified by plants consumption or toxic substance ^(17,36,37). Our results indicated that arbutin (all three concentrations) can increase blood parameters during megavoltage therapeutic x-irradiation on mice. In the groups which were only treated with arbutin there were not any remarkable changes in blood parameters, relative to the control group. Bertrand *et al.* ⁽¹⁷⁾ showed that aged garlic extract as a radioprotector can increase the hematological parameters such as WBC, RBC, LYM, HGB, mean corpuscular volume, and hematocrit value, when injected to rat's blood cells before ionizing radiation. In another study ⁽¹⁵⁾, they showed that lettuce oil could improve all of the above-mentioned blood parameters in rats induced by gamma irradiation. In this case they reported that lettuce oil may reduce the biological hazards of radiation. Also Dong *et al.* ⁽³⁸⁾, expressed that ethanol extract from Ji-Xue-Teng is a strong radioprotective agent due to the recovery from hematopoietic bone marrow damage and oxidative stress of the mice induced by whole body gamma radiation.

Arbutin is a considerable protector (dose

reduction factor = 2.1) and has successfully ameliorated the hematological disturbances induced by radiations which is related to its antioxidant properties ⁽¹⁸⁾, hence, it can sweep free radical, inhibit lipid peroxidation in cell membrane, and stabilize plasma membrane ⁽³⁾.

The effects of the three concentrations of arbutin as a radioprotector were almost similar but a minor difference was observed between them in count of blood cells factors and frequency of micronuclei. The amelioration was more effective in the “2 Gy and 4 Gy x irradiation + 50 mg/kg arbutin” groups in comparison with the “2 Gy and 4 Gy x irradiation + 100 mg/kg arbutin” and “2 Gy and 4 Gy x irradiation + 200 mg/kg arbutin” groups.

As future research, it is suggested that other arbutin concentrations as a radioprotector along with x-radiation in different lengths and doses should be studied to find the precise optimized effect on blood parameters and micronuclei.

CONCLUSION

The micronucleus assay and the evaluation of some blood cell parameters (WBC, CBC, LYM, and HGB) demonstrated that arbutin has radioprotective effects on megavoltage therapeutic X-irradiated mice maybe due to antioxidant and free radical scavenging activities of arbutin. In addition, the study revealed that the radioprotective effect of arbutin was more pronounced with the lower dose (50 mg / kg) compared to the higher ones (100 and 200 mg / kg).

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REFERENCES

1. Sherer MAS, Visconti PJ, Ritenour ER, Haynes K (2017) LIC-Radiation Protection in Medical Radiography: Elsevier Health Sciences.
2. Firouzjah RA, Banaei A, Farhood B, Bakhshandeh M (2019) Dosimetric comparison of four different techniques for supraclavicular irradiation in 3d-conformal radiotherapy of breast cancer. *Health Physics*, **116(5)**: 631-6.
3. Kamran MZ, Ranjan A, Kaur N, Sur S, Tandon V (2016) Radioprotective agents: strategies and translational advances. *Medicinal research reviews*, **36(3)**: 461-93.
4. Giridhar P, Mallick S, Rath GK, Julka PK (2015) Radiation induced lung injury: prediction, assessment and management. *Asian Pac J Cancer Prev*, **16(7)**: 2613-7.
5. Abdi Goushbolagh N, Abedi Firouzjah R, Ebrahimnejad Gorji K, Khosravanipour M, Moradi S, Banaei A, et al. (2018) Estimation of radiation dose-reduction factor for cerium oxide nanoparticles in MRC-5 human lung fibroblastic cells and MCF-7 breast-cancer cells. *Artificial cells, nanomedicine, and biotechnology*, **46(3)**: S1215-S25.
6. Goldstein M and Kastan MB (2015) The DNA damage response: implications for tumor responses to radiation and chemotherapy. *Annual review of medicine*, **66**: 129-43.
7. Hosseini-mehr SJ, Zakaryae V, Froughizadeh M (2006) Oral oxymetholone reduces mortality induced by gamma irradiation in mice through stimulation of hematopoietic cells. *Molecular and cellular biochemistry*, **287(1-2)**: 193-9.
8. Luo L, Wu D, Dai D, Yang Z, Chen L, Liu Q, et al. (2017) Synergistic effects of persistent free radicals and visible radiation on peroxymonosulfate activation by ferric citrate for the decomposition of organic contaminants. *Applied Catalysis B: Environmental*, **205**: 404-11.
9. Adhvaryu M, Srivastav S, Vaniawala S, Reddy M (2008) A comparative study of radioprotection by four Indian medicinal herbs against genotoxicity induced by sub-lethal gamma irradiation in Swiss albino mice. *Int J Radiat Res*, **6(1)**: 19-30.
10. Poprac P, Jomova K, Simunkova M, Kollar V, Rhodes CJ, Valko M (2017) Targeting free radicals in oxidative stress-related human diseases. *Trends in pharmacological sciences*, **38(7)**: 592-607.
11. Jin T, Song T, Deng S, Wang K (2014) Radiation-induced secondary malignancy in prostate cancer: a systematic review and meta-analysis. *Urologia internationalis*, **93(3)**: 279-88.
12. Jamwal VS, Mishra S, Singh A, Kumar R (2014) Free radical scavenging and radioprotective activities of hydroquinone in-vitro. *J Radioprot Res*, **2**: 37-45.
13. Gehlot P and Goyal P (2007) Rectification of radiation-induced damage in swiss albino mice by aloe vera leaf extracts (AVE). *Int J Radiat Res*, **5(2)**: 71-78.
14. Jagetia GC (2007) Radioprotective potential of plants and herbs against the effects of ionizing radiation. *Journal of clinical biochemistry and nutrition*, **40(2)**: 74-81.

15. Abdel-Magied N and Ahmed A (2011) The Protective Role of Lettuce oil (*Lactuca sativa*) against Radiation induced Biological Hazards in Male Rats. *J Rad Res Appl Sci*, **4(3-4)**: 923-38.
16. Targhi RG, Banaei A, Saba V (2019) Radioprotective effect of grape seed extract against gamma irradiation in mouse bone marrow cells. *Journal of cancer research and therapeutics*, **15(3)**: 512.
17. Bertrand KFB, Pascal CDD, Désiré DDP, Odette SN, Myriam M, Sone M, et al. (2016) Effects of aged garlic extract on rat's blood cells parameters after whole-body exposure to ionizing radiation. *American Journal of Pharmacology and Phytotherapy*, **1(1)**: 25-30.
18. Nadi S, Monfared AS, Mozdarani H, Mahmoodzade A, Pouramir M (2016) Effects of arbutin on radiation-induced micronuclei in mice bone marrow cells and its definite dose reduction factor. *Iran J Med Sci*, **41(3)**: 180.
19. Wu L-H, Li P, Zhao Q-L, Piao J-L, Jiao Y-F, Kadowaki M, et al. (2014) Arbutin, an intracellular hydroxyl radical scavenger, protects radiation-induced apoptosis in human lymphoma U937 cells. *Apoptosis*, **19(11)**: 1654-63.
20. Petricic J, Apostolski R, Srepel B (1981) The leaf of the wild pear-tree as an arbutinic herb. *Farmaceutski Vestnik Ljubljana J*, **32**: 107-10.
21. Khanal T, Kim HG, Hwang YP, Kong MJ, Kang MJ, Yeo HK, et al. (2011) Role of metabolism by the human intestinal microflora in arbutin-induced cytotoxicity in HepG2 cell cultures. *Biochemical and biophysical research communications*, **413(2)**: 318-24.
22. Lubsandorzhieva P, Zhigzhitov B, Dargaeva T, Bazarova ZG, Nagaslaeva L (2000) Chromatospectrophotometric determination of arbutin in the leaves of *Bergenia crassifolia* (L.) Fritsch. *Pharmaceutical Chemistry Journal*, **34(5)**: 261-4.
23. Wallner M, Blassnigg S, Marisch K, Pappenheim M, Müllner E, Mölzer C, et al. (2012) Effects of unconjugated bilirubin on chromosomal damage in individuals with Gilberts syndrome measured with the micronucleus cytome assay. *Mutagenesis*, **27(6)**: 731-5.
24. McNamee JP, Flegal FN, Greene HB, Marro L, Wilkins RC (2009) Validation of the cytokinesis-block micronucleus (CBMN) assay for use as a triage biological dosimetry tool. *Radiation protection dosimetry*, **135(4)**: 232-42.
25. Erexson GL, Kligerman AD, Bryant MF, Sontag MR, Halperin EC (1991) Induction of micronuclei by X-radiation in human, mouse and rat peripheral blood lymphocytes. *Mutation Research/Environmental Mutagenesis and Related Subjects*, **253(2)**: 193-8.
26. Benković V, Orsolić N, Knežević AH, Ramić S, Đikić D, Bašić I, et al. (2008) Evaluation of the radioprotective effects of propolis and flavonoids in gamma-irradiated mice: the alkaline comet assay study. *Biological and Pharmaceutical Bulletin*, **31(1)**: 167-72.
27. Karbownik M and Reiter RJ (2000) Antioxidative effects of melatonin in protection against cellular damage caused by ionizing radiation. *Proceedings of the Society for Experimental Biology and Medicine: Minireviews*, **225(1)**: 9-22.
28. Parihar VK, Prabhakar KR, Veerapur VP, Priyadarsini KI, Unnikrishnan MK, Rao CM (2007) Anticlastogenic activity of morin against whole body gamma irradiation in Swiss albino mice. *European Journal of Pharmacology*, **557(1)**: 58-65.
29. Gupta U, Chaudhary R, Goyal P (2010) Post-treatment effects of *Alstonia scholaris* extract against radiation-induced biochemical alterations in Swiss albino mice. *Int J Radiat Res*, **8(3)**: 169.
30. Shaheen A and Hassan S (1991) Radioprotection of whole-body gamma-irradiation-induced alteration in some haematological parameters by cysteine, vitamin E and their combination in rats. *Strahlentherapie und Onkologie: Organ der Deutschen Röntgengesellschaft*, **167(8)**: 498-501.
31. Hosseinimehr S and Nemati A (2006) Radioprotective effects of hesperidin against gamma irradiation in mouse bone marrow cells. *The British journal of radiology*, **79(941)**: 415-8.
32. Mozdarani H and Shahidi M (2003) Potent radioprotective effect of therapeutic doses of ranitidine and famotidine against gamma-rays induced micronuclei *in-vivo*. *Int J Radiat Res*, **1(1)**: 29-35.
33. Shokrzadeh M, Naghshvar F, Ahmadi A, Chabra A, Jeivad F (2014) The potential ameliorative effects of melatonin against cyclophosphamide-induced DNA damage in murine bone marrow cells. *Eur Rev Med Pharmacol Sci*, **18(5)**: 605-11.
34. Hassan I, Chibber S, Naseem I (2010) Ameliorative effect of riboflavin on the cisplatin induced nephrotoxicity and hepatotoxicity under photoillumination. *Food and Chemical Toxicology*, **48(8-9)**: 2052-8.
35. Soyal D, Jindal A, Singh I, Goyal P (2007) Protective capacity of Rosemary extract against radiation induced hepatic injury in mice. *Int J Radiat Res*, **4(4)**: 161-168.
36. Yuan G, Dai S, Yin Z, Lu H, Jia R, Xu J, et al. (2014) Toxicological assessment of combined lead and cadmium: acute and sub-chronic toxicity study in rats. *Food and Chemical Toxicology*, **65**: 260-8.
37. Avci A, Atli T, Ergüder İB, Varlı M, Devrim E, Aras S, et al. (2008) Effects of garlic consumption on plasma and erythrocyte antioxidant parameters in elderly subjects. *Gerontology*, **54(3)**: 173-6.
38. Dong X-Z, Wang Y-N, Tan X, Liu P, Guo D-H, Yan C (2018) Protective Effect of JXT Ethanol Extract on Radiation-Induced Hematopoietic Alteration and Oxidative Stress in the Liver. *Oxidative medicine and cellular longevity*; **2018(6)**: 1-12.