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Development of radiation dosimetric phantoms made from SPC/NaOH/IA-PAE/Rhizophora spp. particleboards

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► ***Original article***

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ABSTRACT

Background: The present study aims to investigate and characterize radiation dosimetric phantom that is made from *Rhizophora spp*. particleboards using SPC/NaOH/IA-PAE adhesive. ***Materials and Methods*:** Modification of soy protein concentrate (SPC) bonded *Rhizophora spp*. particleboard phantoms were carried out using sodium hydroxide (NaOH) with different amount of bio -based cross-linking agent (IA-PAE) with three different categories of particle size. The radiation attenuation parameters were evaluated with photons energies between 16.59 and 25.26 keV derived from X-ray fluorescence. SPC-based plug phantoms were scanned together with the CT density phantom 062M at 80, 120, and 135 kVp, and 250 mAs. ***Results*:** The physical and mechanical properties were found to be improved, while the dimensional stability was decreased through the addition of 15 wt% IA-PAE with decreasing particle size. The lowest HVL and MFP, and the highest μ_m with their corresponding values of Z_{eff} (p-values of 0.054, 0.055, 0.060, and 0.075), and the relatively low X^2 values are the characteristics of samples labeled as A_{15}, B_{15}, C_{10}, and C_{15}. Sample C_{15} gave the closest values of CT number, ED, and DDPs to water for all tested tube voltages. ***Conclusion*:** The results point to the possibility of employing C_{15} as a promising TEPM.

***Keywords*:** *Computed tomography system, RAPs, SPC, rhizophora spp., phantom material.*

INTRODUCTION

In the last few decades, an intense study on the dosimetric properties of various homogeneous phantom materials has been carried out. This was done in the quest of discovering suitable solid tissue-equivalent phantom materials (TEPMs) that would replace water phantom, because of the limitation water poses when applied in conjunction with detectors that are not waterproof. This was in addition to the challenge of localization of the radiation detectors in water [1,2]. However, the use of solid TEPMs is still short of providing good agreement in terms of the radiation attenuation parameters (RAPs) and radiographic properties of water at low and high energies [3]. These properties play a vital role in the field of non-destructive test of medical samples (nuclear medicine and radiology), agricultural

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(characterization of various types of woods and mapping inhomogeneities in different parts of wood), and industrial (to check for voids and insertions in the inner part of the material) interests.

Recently, several comprehensive scientific researchers have been conducted on the suitability of some species of natural mangrove wood as a result of its unique biodegradable, biocompatible, and eco-friendly characteristics [3-6]. *Rhizophora spp.* (*R. spp*.) wood has been in use as a valuable and superior alternative for manufacturing wood composites such as plywood, fiberboards, particleboards, and so on [6]. Subsequently, it has been discovered that it exhibits similarities in dosimetric properties with water and other standard phantom materials at photon energies between 16.59 and 25.26 keV [3, 4, 6]. Consequently, it can be made as an attractive quality materials for different purposes such as diagnostic radiology, nuclear medicine, and other medical applications. Various bio-based and petroleum-based adhesives as widely investigated, have been proffered for *R. spp.* particleboards but with certain disadvantages i.e. stability, firmness, etc. [4-6]. Due to some demerits from petroleum-based adhesive, the development of readily available particleboard phantoms from renewable resources will still be desirable [7].

Moreover, when *R. spp*. composites is reinforced with bio-based adhesive (SPC – Soy protein concentrate), provides outrageous advantages that include flexibility, cost-effectiveness, non-toxicity, conformability, etc., and reduces attendant deleterious environmental effects [6-8]. In spite of this, producing SPC-based particleboards with superior performances in larger quantities remains the main challenge to be addressed on a global scale due to its poor mechanical properties and hydrophilic nature [8, 9]. To enhance these qualities, significant numbers of research has been conducted to modify its molecular structure through chemical cross-linking reactions [5-7, 9, 10]. It was thereafter discovered that there is an increasing demand for itaconic acid polyamidoamine-epichlorohydrin (IA-PAE)/NaOH-based material because of its readily biodegradable and biocompatible nature [7-11]. Therefore, in this study, different concentrations of IA-PAE with NaOH were incorporated into SPC/*R. spp*. with the desire to examine the radiation dosimetric properties which was affected by different sizes of particles.

The present study is a significant one since there is no notable research that has been performed to optimize the properties of *R. spp*. phantoms developed at three different sizes of particles using SPC/NaOH/IA-PAE bio-based adhesive, which is a suitable tissue- and water-equivalent phantom material for both clinical and investigative aspects of radiation therapy dosimetry. The importance of TEPM development is to contribute to the construction of high-quality particleboard phantoms, with comparable dosimetric characteristics to water and other commercial TEPMs, and to avoid the detectable amounts of formaldehyde emissions and other volatile organic compounds using realistic, durable, and inexpensive bio-based materials. This phantom formulation will be useful in the dosimetry of medical health applications.

MATERIALS AND METHODS

Preparation of bio-adhesives

The technique for preparing and synthesizing IA-PAE resin was established in a study previously reported by Samson *et al.* [6] with the characteristics of the resulting product which includes pH 6.68, solid content of 55.96 ± 0.01 wt%, and viscosity of 100.40 ± 0.25 mPa.s. The SPC-based adhesives were prepared at 25°C by dissolving SPC (30 g) under constant stirring in distilled water (70, 65, 60, and 55 g) at 600 rpm for 30 min in accordance with the procedure described by Zhang *et al.* [9]. Subsequently, different concentrations of IA-PAE (0, 5, 10, and 15 wt %) were added to the SPC dispersions and vigorously stirred for 30 min. The result of SPC/IA-PAE slurry mixture was maintained for 1 h at pH 11.0 with 2N NaOH (10 wt %) solution.

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Fabrication of SPC/NaOH/IA-PAE/R. spp. particleboard phantoms

The grinding and segregation of *R. spp.* wood was achieved based on a previous technique detailed by Samson *et al.* [6]. Using an iron-coated cylindrical mould (27 × 0.5 cm^2), the adhesive sample formulations were thoroughly mixed with different *R. spp.* particle sizes and was subsequently cold-compressed at 5 $kgcm^{-2}$, 3 min, and 20°C. The samples were thereafter hot-pressed into particleboards with pressing pressure of 180 $kgcm^{-2}$ and was held for 15 min at 170°C [6]. Just after the mould was released, the phantom sections were smoothed to the desired dimensions and conditioned for two weeks at 20 with relative humidity of 65%. All the measured particleboards were performed in five replicates for each particle size at different concentrations of IA-PAE adhesive, making a total of 75 particleboards manufactured as coded in table 1. The physical, mechanical, and dimensional stability (PMDS) tests were conducted according to JIS A 5908:2003 [12] as well as the technique proposed by Samson *et al.* [6].

Table 1. Comparison of densities of SPC-based R. spp. plug phantoms.

Particle size (µm)	Plug phantoms	Average density (g/cm^3)
149–500	A_0	1.021 ± 0.026
	A_1	1.085 ± 0.007
	A_5	0.979 ± 0.016
	A_{10}	0.997 ± 0.010
	A_{15}	1.010 ± 0.003
74–149	B_0	1.025 ± 0.032
	B_1	1.030 ± 0.015
	B_5	1.034 ± 0.005
	B_{10}	0.994 ± 0.014
	B_{15}	1.009 ± 0.003
≤ 74	C_0	1.097 ± 0.027
	C_1	1.067 ± 0.003
	C_5	1.032 ± 0.011
	C_{10}	0.961 ± 0.012
	C_{15}	1.005 ± 0.002

Plug phantoms measurements for computed tomography (CT) scan

The preparation of SPC-based *R. spp*. plug phantoms compatible with CT electron density phantom (062M, CIRS, Norfolk, USA) was performed through moulding on top of a cylindrical base, 3 cm in diameter and 7 cm in length and was glued vertically into a stack using polyvinyl acetate adhesive. Each sample plug phantom was subsequently trimmed and oriented (figure 1). The results of the selected plug phantoms are listed in table 1.

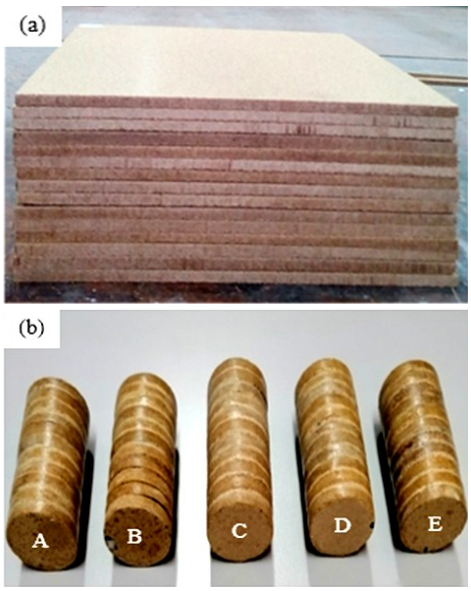


Figure 1. (a) Fabricated particleboards and **(b)** sample plug phantoms. A, B, C, D, and E refers to binderless, 0, 5, 10, and 15 wt% IA-PAE.

Characterization of SPC/NaOH/IA-PAE/R. spp. particleboard phantoms

Thermogravimetric (TG) analysis

TG measurement was carried out on a TG analyser (TGA-7, Norwalk, USA). The average sample size was 5 mg and the nitrogen flow-rate was 20 $mLmin^{-1}$. The temperature range for scanning was from 30-900°C at 10°$Cmin^{-1}$ increment, which was held for 5 min at 900°C.

Fourier transform infrared (FTIR) spectroscopy analysis

FTIR spectra for SPC-based *R. spp*. adhesives were obtained from IRPrestige-21 Shimadzu FTIR spectrometer by diffused reflectance. The samples were prepared by adding about 1 mg of each dried adhesive sample to 100 mg of KBr crystals and compressed at 32 MPa for 3 min to a thickness of 0.5 mm and diameter of 12.71 mm. Measurements were carried out by

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employing 32 scans with a resolution of 4 cm^{-1}, at a spectral wavenumber range of 4000 - 400 cm^{-1}.

Microstructure and energy dispersive X-ray analysis

Samples of size (0.5 × 0.5 × 0.5 cm^3) were mounted on a metal strap holder which were dried in an oven at 120°C, and then gold-coated for imaging at 45 mA for 30 seconds. Images of the coated specimens were observed on the field emission scanning electron microscope (FEI Quanta FEG-650, Netherlands) using 20 kV at 30° with magnifications of 1000×. Analyses were performed under vacuum conditions. The effective atomic number (Z_{eff}) and electron fraction (α_i) of the samples were determined using Equation 1 [6, 13].

$$Z_{eff} = \frac{\sigma_a}{\sigma_e} = \left[\sum_i^r \alpha_i {z_i}^m\right]^{1/m} = \left[\sum_i^r \frac{N_g^i}{N_g} {z_i}^m\right]^{1/m} = \left[\sum_i^r \frac{{\omega_i z_i}/{A_i}}{\sum_i \left({\omega_i z_i}/{A_i}\right)} {z_i}^m\right]^{1/m} \quad (1)$$

where, σ_a,σ_a,z_i,α_i,A_i, and ω_i are the atomic cross-section, electronic cross-section, atomic number, electron fraction, effective atomic weight, and fractional weight of i^{th} element (i=1,2,3,..,r) in the sample materials, while m is the experimental coefficient for biological materials and water with a value of 3.4 [13].

X-ray diffraction (XRD) analysis

The crystallinity of the samples was investigated by utilizing a Bruker D8 Advance X-ray diffractometer operating with Cu-Kα1 X-ray radiation of 1.5406 Å at 40 kV_p and 40 mA, using a cobalt source detector and a 2θ scan ranging from 5° - 80° at 2°/min. Structural analysis of the XRD data was performed via EVA 5+ software. The determination of the index of crystallinity was achieved by equation 2 [6].

$$C_rI(\%) = \left[\frac{I_{200} - I_{am}}{I_{200}}\right] \times 100 \quad (2)$$

where, I_{200} and I_{am} are the peak intensities corresponding to crystalline and amorphous fractions.

Particleboards evaluation

RAPs

The photon attenuation measurements were performed using a LEGe detector system combined with a multichannel analyzer (MCA-3 series), high voltage source with positive polarity, absorber particleboard samples, a radioactive point source, and Pb collimator. The detector had an energy resolution of 400 eV for 59.54 keV photons from ^{241}Am with an active crystal area of 2 cm^2 and a diameter of 1.6 cm, kept at liquid nitrogen temperature (-196°C). Both distances from the samples to the metal plate and the detector were 7 cm and 8.9 cm, while the diameter of the Pb collimator was 0.3 cm. Particleboard samples were prepared and irradiated according to the technique proposed by Samson *et al.* [6]. The mass attenuation coefficient (μ_m) being a function of only energy and the atomic number of the material is calculated using equation 3:

$$\mu_m = \frac{1}{x.\rho} \ln\left(\frac{I_o}{I}\right) \quad (3)$$

where, I and I_o are the number of counts under the photons peak with and without samples. x and ρ are the physical thickness and density. In the case of a compound, the μ_m of any composite material consisting of several mixture of chemical elements present in the sample is measured as given in equation 4:

$$\mu_m = \left(\frac{\mu}{\rho}\right)_{composite} = \omega_1\left(\frac{\mu}{\rho}\right)_1 + \omega_2\left(\frac{\mu}{\rho}\right)_2 + \cdots = \sum_{i=1}^{N} \omega_i \left(\frac{\mu}{\rho}\right)_i \quad (4)$$

where, ω_i and $(\mu/\rho)_i$ denote the weight fraction and μ_m of the constituent elements i^{th}.

The half-value layer (HVL) is the thickness of a specified material used in dividing the photon intensity and assess the capacity of radiation attenuation for any material, equation 5 provides the expression for the HVL value [1,14].

$$HVL\ (X_{1/2}) = \frac{\ln 2}{(\mu_m \times \rho)} \quad (5)$$

The mean free path (MFP) provides an

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approximation of the average distance travelled by the photon in the sample material before it can be absorbed or scattered [14]. It can be calculated as shown in equation 6:

$$MFP\ (\lambda) = \frac{1}{(\mu_m \times \rho)} \quad (6)$$

Equation 7 is used for the calculation of associated standard error in the experimental μ_m values.

$$\frac{\Delta\mu_m}{\mu_m} = \sqrt{\left(\frac{\Delta I_o}{I_o \mu x \rho}\right)^2 + \left(\frac{\Delta I}{I \mu x \rho}\right)^2 + \left(\frac{\Delta x.\rho}{x.\rho}\right)^2} \quad (7)$$

where, the first, second, and third term denotes the statistical fluctuation intensities of the transmitted and incident photon beam and its uncertainty in measuring sample mass-thickness. All data were statistically analyzed using SPSS V22.0 and the results was compared with the calculated values of water (XCOM) [15].

Measurement of CT numbers, electron densities (ED) and density distribution profiles (DDPs)

CT scan was performed at three X-ray CT tube voltages (80, 120, and 135 kVp), current at 250 mAs, in-plane resolution of 0.714 × 0.714 mm²/pixel, and slice thickness of 1 mm using Somatom Definition. AS+ scanner. Slice acquisition times were 4 seconds per scan and 365.625 mm field of reconstruction was captured in a 512 pixel × 512 pixel image. The CT number was measured using equation 8 [16].

$$\text{CT number} = K\left(\frac{\mu - \mu_w}{\mu_w}\right) \quad (8)$$

where, μ and μ_w are the linear attenuation coefficients of the sample and water, and K is the magnification contrast with the value of 1000. The discrepancy in each scanned sample from the computed value of water was calculated using the Chi-square (x^2) goodness of fit test shown in equation 9:

$$\chi^2 = \sum_{i=1}^{N}\left(\frac{\text{CT}_{w_i} - f(\text{CT}_i)}{\sigma_i}\right)^2 \quad (9)$$

where, CT_{wi} and CT_i are the CT number of water and sample, and σ is the uncertainty in the sample plug phantoms. A couple of standard TEMs with known densities were scanned as presented in figure 2, since CT numbers varies among scanners from different manufacturers, to establish the linear correlation of the calibration curve.

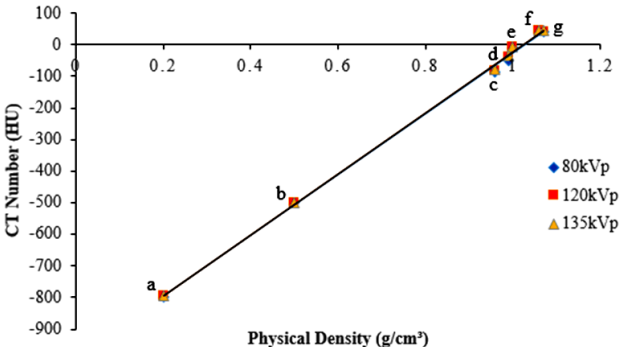


Figure 2. Calibration curve between CT numbers and physical densities. **(a)** lung (inhale), **(b)** lung (exhale), **(c)** adipose, **(d)** breast (50/50), **(e)** water, **(f)** muscle, and **(g)** liver. (Error bars cannot be seen because they are smaller than the symbols).

RESULTS

TG properties

The thermal stability study showed that TG and DTG techniques permitted the specification of the temperature range, whereby the samples were thermally stable within 0°C-280.67°C (figure 3). The ratio of weight loss was found to be within 4.51-11.66% at the first peak, while the second region was initiated in the range between 71.48-87.57% (figure 3(a)). These stages were assigned to the evaporation of residual moisture, degradation of the main skeleton structure, and disruption of the quaternary structure accompanied by partial protein denaturation, which is involved with the thermal breakage of C-C, C-N, and C-O bonds [6,17]. SPC pyrolysis was seen to peak between 180-400°C, while the DTG curve appears to be range of 400-900°C (figure (3b)). These findings confirm an improvement in the thermal stability following NaOH/IA-PAE incorporation that suggest the presence of a chemical cross-linking reaction.

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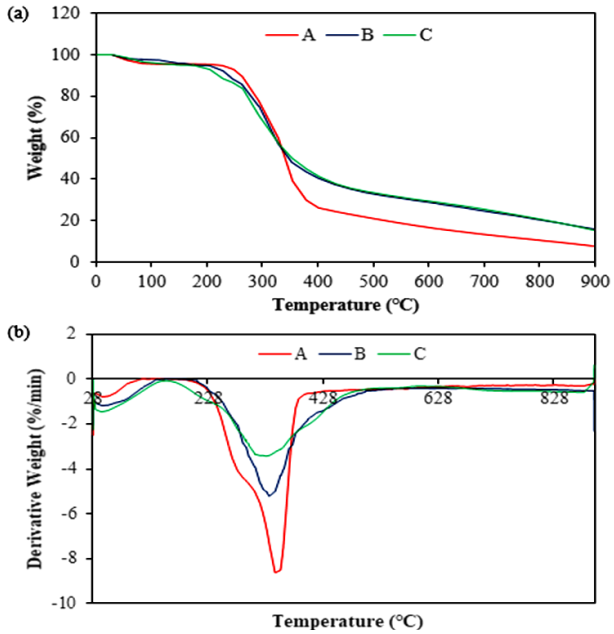


Figure 3. TG properties: **(a)** TG and **(b)** DTG curves. ABinderless R. spp., BSPC/R. spp., and CSPC/NaOH/IA-PAE/R. spp.

FTIR analysis

The characteristic broad absorption peaks were observed at 3000-3442.94 cm^{-1}, which are attributable to the free and bonded O-H and N-H groups (figure 4). The bands relating to C=O stretching (1658.78 cm^{-1}), N-H bending (1575.84 cm^{-1}), C-N stretching/N-H bending vibrations (1317.38-1259.52 cm^{-1}) were ascribed to amide I, amide II, and amide III [6]. In addition, the bands at 1157.29, 1504.48, 1456.26, 1051.20, and 2376.30-2341.58 cm^{-1} were assigned to ester C-O moieties, C-H deformation of aliphatic, skeletal vibration of C=C, stretching vibration of COO-/C-O, and amino hydrohalides [11,17].

Microscopy images

Figure 5(a) exhibits the stacks of the binderless R. spp., showing some number of gaps and voids in the composites, which could weaken the bonding between fiber and matrix (3). In figure 5(b) it is visible that some agglomeration occurred in the composite particleboards. This is due to the poor interfacial adhesion with the matrix, thus the composite toughness was increased. Figure 5(c-d) depicts that the NaOH/IA-PAE (15 wt%) distorted the void spaces, align uneven fibers, and bring about compact fracture surfaces within the particleboards (9,10). The computed Z_{eff} values for the selected samples lies within the range of 7.157.59 (table 2).

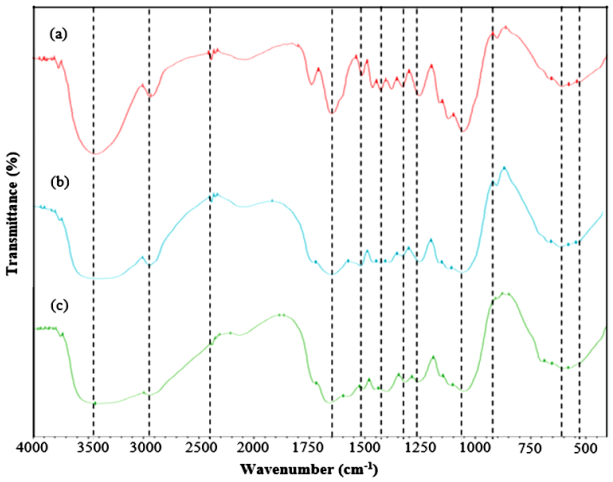


Figure 4. FTIR spectra: **(a)** Binderless R. spp., **(b)** SPC/R. spp., and **(c)** SPC/NaOH/IA-PAE/R. spp.

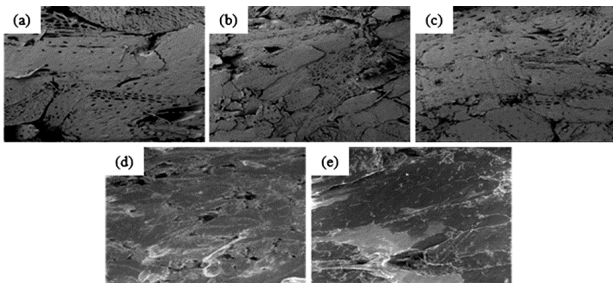


Figure 5. SEM images: **(a)** Binderless R. spp., **(b)** untreated SPC/R. spp., **(c)** SPC/NaOH/IA-PAE/R. spp. (5 wt%), **(d)** SPC/NaOH/IA-PAE/R. spp. (10 wt%), and (e) SPC/NaOH/IA-PAE/R. spp. (15 wt%).

Structural Properties

The results of the XRD measurement are displayed in figure 6. The diffraction patterns observed at 2θ with the d-spacing were estimated at 14.28° (6.165 Å), 22.41° (3.998 Å), and 34.75° (2.582 Å) for SPC/NaOH/IA-PAE/*R. spp.*, 57.4³∫° (⁰.⁰49 Å), 66.6³ ° (7.³97 Å), 78.8² ° (2.600 Å) for SPC/NaOH/*R. spp.*, 13.73° (6.405 Å), 22.12° (3.954 Å), 34.52° (2.596 Å) for SPC/*R. spp.*, and 58.⁰¹ ° (⁰.475 Å), 66.7¹ ° (7.³⁰4 Å), and 34.52° (2.595 Å) for binderless *R. spp.*, particleboards, matched to the (101), (002), and (004) planes. The overall results for the crystallinity index are 42.27%, 32.12%, 27.98%, and 41.58%, respectively.

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Table 2. Elemental contents and Z_{eff} values.

Sample	Elemental composition (%)												Z_{eff}
	H	C	N	O	Na	Mg	P	S	Cl	K	Ca	Zn	
Water	11.11	-	-	88.89	-	-	-	-	-	-	-	-	7.50[a]
Perspex	31.97	59.98	-	8.05	-	-	-	-	-	-	-	-	5.21[a]
C_0	-	49.04	1.80	48.92	-	-	-	0.24	-	-	-	-	7.15[b]
C_1	-	49.33	4.42	45.21	0.34	-	-	-	0.32	0.16	0.22	-	7.40[b]
C_5	-	47.96	5.17	45.81	0.33	0.09	-	-	0.41	0.09	0.14	-	7.41[b]
C_{10}	-	51.24	4.54	43.04	0.22	-	0.10	0.10	0.35	0.20	0.17	-	7.45[b]
C_{15}	-	49.73	4.88	44.35	0.25	0.10	-	-	0.27	0.25	0.15	0.02	7.59[b]
Almond gum	6.03	41.72	0.27	51.98	-	-	-	-	-	-	-	-	7.15[c]

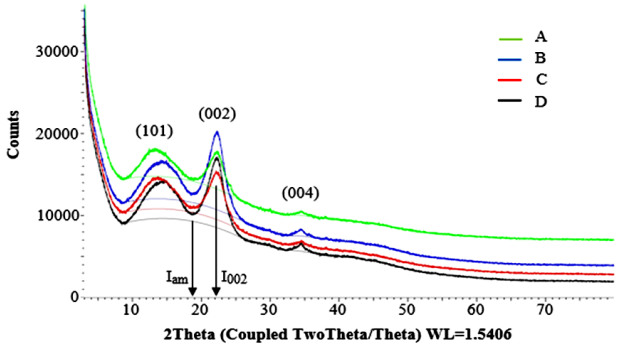


Figure 6. Indexed XRD patterns: ASPC/NaOH/IA-PAE/R. spp., BSPC/NaOH/R. spp., CSPC/R. spp., and DBinderless R. spp. particleboards.

The results show remarkable values of MC and SC, ranging from 6.5 ± 0.2-8.4 ± 0.3% and 29.5 ± 0.2-39.1 ± 0.9% (table 3). The maxima modulus of rupture (MOR) and elasticity (MOE), and internal bonding (IB) values were ≤ 74 µm, while the minima values appear at 74-149 µm and 149-500 µm. Meanwhile, samples of about 15 wt% of IA-PAE at ≤ 74 µm exhibited a decrease in water absorption (WA) and thickness swelling (TS) ratio [12]. On the other hand, the higher WA and TS observed with 0, 5, and 10 wt% of IA-PAE, might be due to the elimination of hydrophobic substances, which promote water penetration and retention [6,7].

Table 3. Comparison of PMDS properties.

Particle size (µm)	Sample	MC (%)	SC (%)	IB (MPa)	MOR (MPa)	MOE (GPa)	WA (%)	TS (%)
149–500	A_0	7.9 ± 0.2	29.5 ± 0.2	0.08 ± 0.01	2.3 ± 0.14	0.34 ± 0.09	87.4 ± 0.2	69.9 ± 0.3
	A_1	7.2 ± 0.4	31.7 ± 0.7	0.15 ± 0.02	3.3 ± 0.16	0.98 ± 0.05	79.5 ± 0.5	66.4 ± 0.2
	A_5	7.9 ± 0.1	32.9 ± 0.3	0.31 ± 0.02	10.5 ± 0.23	1.12 ± 0.09	44.3 ± 0.1	55.4 ± 0.1
	A_{10}	6.8 ± 0.1	33.0 ± 0.2	0.39 ± 0.08	11.8 ± 0.07	2.29 ± 0.03	33.7 ± 0.5	48.5 ± 0.3
	A_{15}	8.2 ± 0.4	34.9 ± 0.4	0.42 ± 0.05	13.1 ± 0.39	3.99 ± 0.07	32.5 ± 0.6	36.9 ± 0.1
74–149	B_0	6.9 ± 0.4	30.9 ± 0.1	0.09 ± 0.02	3.9 ± 0.07	0.47 ± 0.09	77.1 ± 0.4	66.3 ± 0.2
	B_1	6.5 ± 0.2	33.0 ± 0.4	0.11 ± 0.08	5.5 ± 0.15	1.15 ± 0.27	72.8 ± 0.1	62.9 ± 0.1
	B_5	7.9 ± 0.3	33.2 ± 0.4	0.30 ± 0.03	10.4 ± 0.10	2.56 ± 0.86	52.5 ± 0.2	45.4 ± 0.1
	B_{10}	6.7 ± 0.2	34.7 ± 0.2	0.44 ± 0.09	13.5 ± 0.08	5.25 ± 0.22	47.5 ± 0.1	31.4 ± 0.3
	B_{15}	6.8 ± 0.2	35.7 ± 0.6	0.48 ± 0.02	16.2 ± 0.17	6.05 ± 0.78	45.9 ± 0.2	28.1 ± 0.2
≤ 74	C_0	7.5 ± 0.7	32.6 ± 0.4	0.08 ± 0.06	3.9 ± 0.74	1.35 ± 0.17	71.1 ± 0.4	60.1 ± 0.9
	C_1	8.4 ± 0.3	31.1 ± 0.9	0.28 ± 0.04	4.0 ± 0.60	1.81 ± 0.33	52.1 ± 0.2	59.5 ± 0.9
	C_5	8.2 ± 0.2	34.6 ± 0.2	0.52 ± 0.01	10.8 ± 0.37	4.40 ± 1.03	26.6 ± 0.6	22.3 ± 0.6
	C_{10}	7.5 ± 0.3	36.2 ± 0.1	0.54 ± 0.02	14.2 ± 0.23	6.28 ± 1.15	19.6 ± 0.3	14.9 ± 0.9
	C_{15}	7.2 ± 0.6	39.1 ± 0.9	0.56 ± 0.01	18.4 ± 0.28	7.58 ± 1.83	18.1 ± 0.9	11.1 ± 0.7

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Evaluation of RAPs

From figure 7, the variation of μ_m values with that of water (XCOM) have similar characteristics for the photon energies at the three selected particle sizes, which satisfactorily matches the results of previous studies [3,6]. As the sample particle sizes decrease and IA-PAE increases, the μ_m values vary from 1.12-0.39 cm^2g^{-1}. Samples A_{15}, B_{15}, C_{10}, and C_{15} with p-values of 0.054, 0.055, 0.060, and 0.075 show no significant difference to that of water as presented in table 4. Therefore, it can easily be seen that HVL and MFP values are minima at lower energies and increases with an increase in photon energy (figures 8 and 9). These results indicated that samples A_{15}, B_{15}, C_{10}, and C_{15} can provide appropriate features as suitable TEPMs.

Evaluation of CT numbers, ED, and DDPs

From table 5, C_{15} has the closest mean CT numbers and ED values, as well as DDPs (figure 10) to water. This is closely followed by B_{15}, A_{15}, and C_{10} at the three X-ray CT tube voltages depicted by the x^2 values (table 6), which revealed the nearest value of C_{15} to water with insignificant difference.

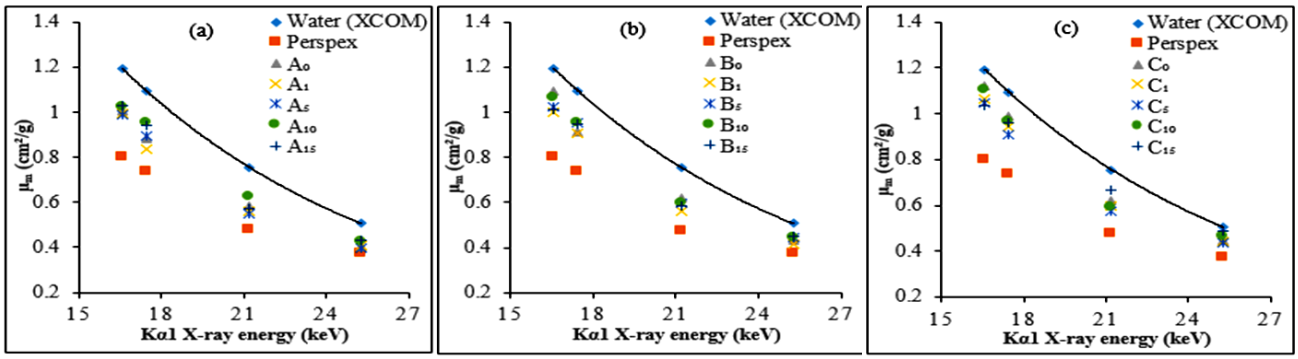


Figure 7. Variation of: **(a)** 149500, **(b)** 74149, and **(c)** 74 µm. (Error bars cannot be seen because they are smaller than the symbols).

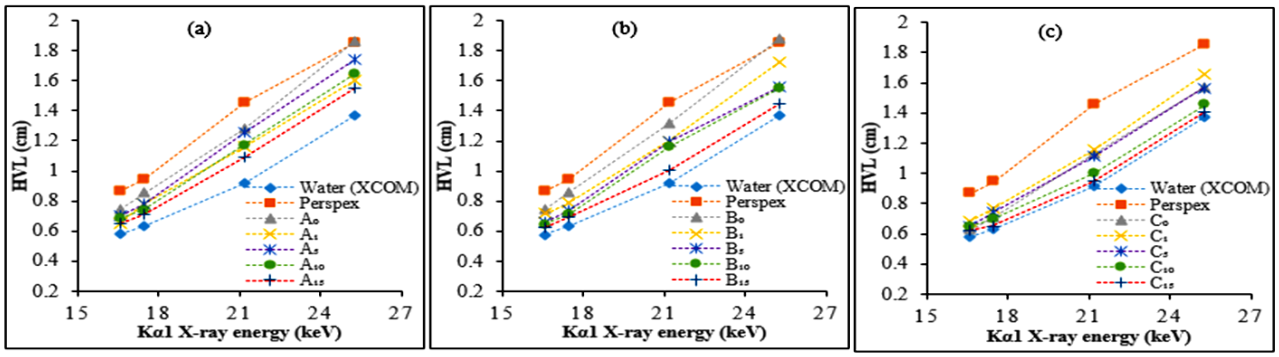


Figure 8. HVL: **(a)** 149500, **(b)** 74149, and **(c)** 74 µm. (Error bars cannot be seen because they are smaller than the symbols).

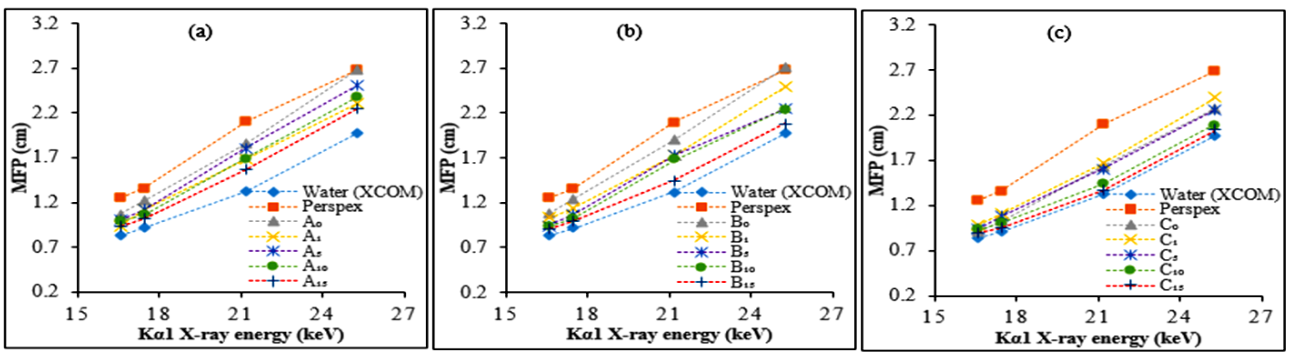


Figure 9. MFP: **(a)** 149500, **(b)** 74149, and **(c)** 74 µm. (Error bars cannot be seen because they are smaller than the symbols).

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Table 4. Paired t-test of the of particleboard phantoms compared to water (XCOM).

Pair	Paired Differences					t	df	Sig. (2-tailed)
	Mean (d)	Std. Dev. (σ_d)	Std. Error Mean	95% Confidence Interval of the difference				
				Lower	Upper			
A_0	0.168	0.047	0.023	0.0434	0.1921	5.043	3	0.015
A_1	0.185	0.067	0.034	0.0479	0.2627	4.600	3	0.019
A_5	0.180	0.049	0.024	0.0325	0.1875	4.519	3	0.020
A_{10}	0.130	0.038	0.019	0.0102	0.1298	3.723	3	0.034
A_{15}	0.145	0.048	0.024	-0.0023	0.1513	3.087	3	0.054
B_0	0.121	0.046	0.028	0.0246	0.1764	4.216	3	0.021
B_1	0.167	0.049	0.025	0.0373	0.1962	4.677	3	0.018
B_5	0.140	0.034	0.017	0.0153	0.1247	4.072	3	0.027
B_{10}	0.122	0.042	0.021	0.0054	0.1391	3.441	3	0.041
B_{15}	0.139	0.058	0.029	-0.0035	0.1805	3.061	3	0.055
C_0	0.095	0.034	0.017	0.0350	0.1439	5.229	3	0.014
C_1	0.127	0.042	0.021	0.0107	0.1428	3.698	3	0.034
C_5	0.154	0.053	0.027	0.0043	0.1743	3.342	3	0.044
C_{10}	0.117	0.035	0.018	-0.0039	0.1084	2.960	3	0.060
C_{15}	0.142	0.068	0.034	-0.0169	0.1999	2.687	3	0.075

Table 5. Comparison of computed CT number and ED.

Sample	80 kVp			120 kVp			135 kVp		
	Average CT No	Average SD	ED x 10^{23} (elect/cm^3)	Average CT No	Average SD	ED x10^{23} (elect/cm^3)	Average CT No	Average SD	ED x 10^{23} (elect/cm^3)
A_0	84.334	29.271	3.669	75.928	20.267	3.662	84.687	20.693	3.662
B_0	3.852	41.688	3.388	1.705	28.613	3.379	1.298	27.813	3.369
C_0	71.513	38.701	3.625	46.719	24.418	3.551	65.589	22.931	3.595
A_1	55.520	32.284	3.569	94.422	24.421	3.732	97.725	21.925	3.707
B_1	7.268	34.311	3.400	1.515	19.478	3.379	3.287	19.4134	3.377
C_1	42.279	35.844	3.523	49.686	19.374	3.562	35.105	20.199	3.488
A_5	-44.062	26.463	3.220	59.753	26.207	3.600	-55.553	17.221	3.171
B_5	10.471	36.999	3.411	16.083	24.935	3.434	15.966	23.640	3.421
C_5	8.393	33.509	3.404	-43.559	21.678	3.208	-34.087	19.619	3.246
A_{10}	-27.963	29.995	3.277	-55.686	15.968	3.162	-41.338	21.928	3.221
B_{10}	-27.239	38.760	3.279	-33.666	25.953	3.245	-32.728	25.229	3.251
C_{10}	-59.120	36.027	3.168	-70.707	26.663	3.105	-53.909	24.853	3.177
A_{15}	-16.841	28.099	3.316	-10.077	16.941	3.335	-12.791	20.409	3.321
B_{15}	-13.211	36.525	3.328	-18.011	21.267	3.305	-18.522	21.609	3.301
C_{15}	-13.691	35.477	3.327	-9.669	20.220	3.336	-5.993	19.631	3.344
Water	-6.162	31.663	3.34	-2.338	15.147	3.34	-0.975	14.691	3.34

Table 6. x^2 for CT numbers.

Energy (kVp)	x^2 to water														
	149-500 µm					74-149 µm					≤ 74 µm				
	A_0	A_1	A_5	A_{10}	A_{15}	B_0	B_1	B_5	B_{10}	B_{15}	C_0	C_1	C_5	C_{10}	C_{15}
80	0.95	1.03	0.75	0.31	0.25	0.49	0.55	0.29	0.20	0.12	0.31	0.92	0.67	0.29	0.05
120	1.07	0.60	0.53	0.39	0.19	0.64	0.31	0.41	0.50	0.18	0.36	0.45	0.09	0.08	0.06
135	0.69	0.47	0.27	0.10	0.08	0.53	0.71	0.33	0.06	0.05	0.52	0.88	0.22	0.09	0.04
Total x^2	2.72	2.11	1.55	0.80	0.52	1.66	1.57	1.03	0.77	0.35	1.18	2.25	0.99	0.47	0.15

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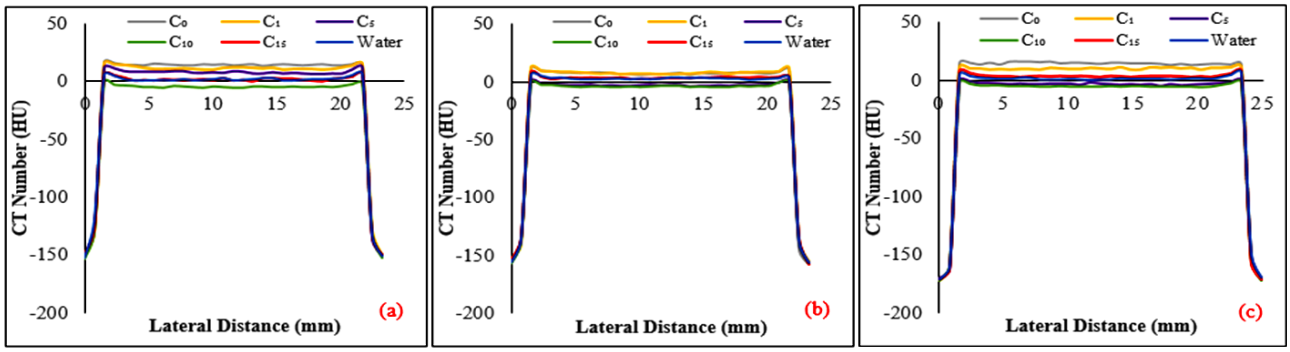


Figure 10. DDPs: **(a)** 80 kVp, **(b)** 120 kVp, and **(c)** 135 kVp.

DISCUSSION

As it can be seen, the difference between the primary and secondary structures of SPC does not significantly affect the thermal degradation process in a nitrogen atmosphere. The main reason for these observations can be explained through figure 3(a-b), which the denaturation of SPC promotes the intra- and inter-molecular cross-linking of amino acid residues, as well as the formation of disulfide cross-links and hydrophobic bonds [6].

The findings confirm the spectra of figure 4 (a), figure 4(b), and figure 4(c) absorption peak regions of the hydroxyl groups to be increased in the order of figure 4(c) < figure 4(b) < figure 4 (a) after treatment with IA-PAE owing to the high protein content in figure 4(c) [11]. It was revealed that in SPC/NaOH/IA-PAE spectra, all essential structural absorption bands of SPC still demonstrated the absence of apparent chemical reaction between the adhesives.

It is worth pointing out that the sample of about 15 wt% IA-PAE and ≤ 74 μm particle size are distributed much more homogeneously compared with the other selected samples. Furthermore, the maxima Z_{eff} were observed for C_{15} with nearly the exact value to water. This shows that the presence of NaOH/IA-PAE adhesive breeds a uniform blend of SPC/*R. spp*. compressed cell walls and ground parenchymatous tissues.

Herein, the studies show, SPC/*R. spp.* particleboard has an amorphous structure, while SPC/NaOH/IA-PAE/*R. spp*. exhibit crystalline structure as the highest crystallinity index with enhanced (101) and (002) peak intensity due to the increasing IA-PAE concentrations. This result is consistent with the result already obtained by other researchers [6, 7, 9, 10, 17].

It is confirmed that the changes produced agrees toward the adhesion by mechanical anchorage, resulting in excellent structural stability, especially for the SPC/NaOH/*R. spp*./IA -PAE (15 wt%) [6, 8]. Moreover, at specific IA-PAE content, the WA and TS have been reduced by decreasing the particle size which is particularly important since such material, when used as a phantom, could be more resistant in the wet state.

Evidently, RAPs are affected by the chemical contents of the particleboard samples as well as incident photon energy; as the photoelectric effect plays a dominant role. Samples A_{15}, B_{15}, C_{10}, and C_{15} exhibited the highest values of μ_m, while the HVL and MFP values of these particleboards were the lowest. This can be attributed to the more significant presence of high atomic number elements and thus provides greater radiation absorption ability.

It is apparent that the C_{15} exhibits substantially matchless performance due to the compact homogeneous surface and close interaction. In addition, higher X-ray CT tube voltage provides good density homogeneity and convenient DDPs to water as indicated by the lower relative standard deviation. The results achieved revealed the possibility of employing C_{15} as a reference in the development of TEPM for use in CT imaging studies.

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CONCLUSION

In conclusion, SPC/NaOH/IA-PAE/*R spp*. particleboard phantom with 15 wt% increment in IA-PAE addition level and particle size of ≤ 74 µm, show optimum qualities among the selected samples and was observed to be ideal for utilization in radiation dosimetry as TEPM.

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Conflicts of interest*: *Declared none.

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